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Mandible strike kinematics of the trap-jaw ant genus *Anochetus* Mayr (Hymenoptera: Formicidae)

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8	Mandible Strike Kinematics of the Trap-Jaw Ant Genus Anochetus Mayr (Hymenoptera:
9	Formicidae)
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- Abstract: High-speed power-amplification mechanisms are common throughout the animal kingdom. In ants, power-amplified trap-jaw mandibles have evolved independently at least four times, including once in the subfamily Ponerinae which contains the sister genera *Odontomachus* and *Anochetus*. In *Odontomachus*, mandible strikes have been relatively well described and can occur in less than 0.15ms and reach speeds of over 60m/s. In contrast, the kinematics of
- 6 mandible strikes have not been examined in *Anochetus*, whose species are smaller and
- 7 morphologically distinct from *Odontomachus*. In this study, we describe the mandible strike
- 8 kinematics of four species of *Anochetus* representative of the morphological, phylogenetic, and
- 9 size diversity present within the genus. We also compare their strikes to two representative
- species of *Odontomachus*. We found that two species, *Anochetus targionii* and *Anochetus*
- 11 paripungens, have mandible strikes that overall closely resemble those found in Odontomachus,
- reaching a mean maximum rotational velocity and acceleration of around $3.7x10^4$ rad/s and
- $8.5 \times 10^8 \text{ rad/s}^2$, respectively. This performance is consistent with predictions based on body size
- scaling relationships described for *Odontomachus*. In contrast, *Anochetus horridus* and
- Anochetus emarginatus have slower strikes relative to the other species of Anochetus and
- 16 Odontomachus, reaching mean maximum rotational velocity and acceleration of around 1.3x10⁴
- rad/s and, $2x10^8$ rad/s², respectively. This variation in strike performance among species of
- 18 Anochetus likely reflects differences in evolutionary history, physiology, and natural history
- 19 among species.

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21 Keywords: Comparative biomechanics, catapult mechanism, functional morphology, power-

22 amplification, mandible strike, Formicidae, kinematics

Introduction

- 25 Speed is an important trait for the success of many animals. The ability to move quickly to evade
- 26 predators or capture prey is a significant selection pressure. How fast animals can move depends,
- 27 in part, on how quickly they can contract muscles to accelerate their limbs. For small animals
- such as arthropods, the distance they move their limbs is short, requiring higher acceleration
- values to obtain peak velocities similar to a larger animal (Alexander, 1988). Additionally,
- muscle fibers are limited by how quickly they can contract (Ellington, 1985; Gronenberg, 1996a;
- 31 Askew & Marsh, 2002), reducing the ability of organisms to achieve high accelerations via direct

- muscle contraction. To overcome these constraints, many animals have evolved catapult
 mechanisms that use a combination of springs, levers, and latches to store and transfer elastic
- and mechanical energy, allowing them to accelerate their limbs more quickly than possible
- 4 through muscle contraction alone (Gronenberg, 1996a; Patek et al., 2015). These power
- 5 amplification or impulse mechanisms have evolved repeatedly in a diverse range of taxa, and
- 6 result in some of the fastest movements known in the animal kingdom (Gronenberg, 1996a;
- 7 Patek et al., 2015).
- 8 In ants, power-amplifying "trap-jaw" mandibles have evolved independently for prey
- 9 capture at least four times across three subfamilies (Larabee & Suarez, 2014) and possibly
- repeatedly in a single myrmicine genus (Ward et al., 2014). While each group has different
- morphological adaptations to store and amplify energy, most have structures that perform similar
- functions (Gronenberg et al., 1993; Gronenberg & Tautz, 1994; Gronenberg, 1996b; Gronenberg
- 2017). & Ehmer, 1996; Gronenberg *et al.*, 1998; Just & Gronenberg, 1999; Larabee *et al.*, 2017).
- Elongated mandibles are locked open with a latch mechanism, allowing elastic strain energy to
- be stored within the head capsule as enlarged mandible closer (adductor) muscles contract. When
- inter-mandibular mechanosensory trigger hairs are stimulated, the latch is released by the
- contraction of fast-twitch muscles, allowing the mandibles to close extremely rapidly (Patek. et
- 18 *al.*, 2006; Spagna *et al.*, 2008; Larabee *et al.*, 2017).
- Most research on the functional morphology and strike kinematics of trap-jaw ants has
- focused on the genus *Odontomachus* Latreille. In this genus, the mandibles are held open by a
- latch formed through interactions between the basal condyle of the mandibles and the mandible
- sockets (Gronenberg et al., 1993). Elastic energy for the mandible strike is likely stored, in part,
- within enlarged mandibular apodemes, although the exact location of energy storage remains
- unknown (Gronenberg et al., 1993). Strikes are stimulated by contact with trigger hairs on the
- 25 mandibles that connect to "giant" sensory neurons (Gronenberg & Tautz, 1994). During a strike,
- 26 Odontomachus mandibles reach peak maximum velocities between 35 and 67 m/s, accelerate
- tens to hundreds of thousands times faster than gravity, and generate forces over 300 times the
- mass of an individual ant (Patek et al., 2006; Spagna et al., 2008). Due to the high force-to-body
- 29 mass ratio of these mechanisms, some *Odontomachus* species use their mandibles to repel small
- 30 intruders out of their nests (Carlin & Gladstein, 1989), and to perform horizontal or vertical

jumps, which can increase survivorship during encounters with predators such as antlions (Patek *et al.*, 2006; Spagna *et al.*, 2009; Larabee & Suarez, 2015).

Anochetus Mayr is the sister genus to Odontomachus and together they represent a single evolutionary transition to power amplified mandibles within the subfamily Ponerinae (Larabee et al., 2016). Relative to Odontomachus, ants in the genus Anochetus are generally smaller, have a distinct head morphology, and frequently ambush and sting their prey when hunting. In contrast, Odontomachus species typically rely on the crushing force of their mandibles alone for prey capture (Brown, 1976; Brown, 1978; De la Mora et al., 2008). Performance of mandible strikes is correlated with body size in Odontomachus (Spagna et al., 2008), with smaller species generally producing faster but less forceful strikes. While the performance properties of Anochetus mandible strikes have never been quantified, they are likely to have similar scaling relationships between performance and body size to species in the genus Odontomachus.

In this study, we describe the mandible strike kinematics of four species of *Anochetus*. In addition to quantifying interspecific variation within *Anochetus*, we compare their strike performance to two species of *Odontomachus*. While *Odontomachus* are generally larger than *Anochetus*, one of our species (*A. emarginatus*) is similar in size to smaller *Odontomachus* (Larabee *et al.*, 2016). We use these data to test the following hypotheses based on the scaling of body size with performance seen in *Odontomachus*: 1) *Anochetus* mandibles will achieve higher accelerations and velocities, and 2) *Anochetus* mandibles will generate less kinetic energy relative to larger species of *Odontomachus*. We also used x-ray micro-tomography (microCT), a method of visualizing three dimensional morphological structures (Friedrich *et al.*, 2014; Wipfler *et al.*, 2016) to characterize the internal morphology of *Anochetus*'s trap-jaw mechanism.

Materials and Methods

Colony Collection and Maintenance

We selected four species of *Anochetus* representative of the morphological, phylogenetic and size diversity within the genus (Figure 1): two species, *A. horridus* and *A. emarginatus*, from the *emarginatus* species group (Brown, 1976; Brown, 1978; clade F in Larabee *et al.*, 2016); one species, *Anochetus targionii*, from the *mayri* species group (Brown, 1976: Brown, 1978; clade F in Larabee *et al.*, 2016); and one species, *Anochetus paripungens*, from the *rectangularis* species group (Brown, 1976; Brown, 1978; clade G in Larabee *et al.*, 2016). We compared the

- 1 kinematics of *Anochetus* to two species of *Odontomachus* chosen to represent the large variation
- 2 in size seen in this genus Odontomachus chelifer (the largest; Figure 1 E) and O. ruginodis (one
- 3 of the smallest; Figure 1 F). The kinematics of O. ruginodis mandible strikes were originally
- 4 described in Spagna et al. (2008); videos recorded for that study were reanalyzed using the
- 5 methods described below.

7 Filming and Video Analysis

- 8 We filmed mandible strikes with a Phantom V9.1 high speed camera (Vision Research Co.,
- 9 Wayne, NJ) connected to a Zeiss SteREO Discovery V20 Microscope (Carl Zeiss Inc.,
- Oberkochen, Germany). Three individuals of A. horridus were filmed using a Phantom Miro eX4
- camera (Vision Research Co., Wayne, NJ) instead of the V9.1. Individual ants were restrained by
- attaching a size three insect pin (Bioquip, Rancho Dominguez CA) to the vertex of their head
- with utility wax (Kerr Laboratory Products, Orange, CA). This pin was placed in a custom
- micromanipulator to position the ant's mandibles into frame prior to striking. Strikes were
- stimulated by either blowing on the ant or touching the trigger hairs / antennae with an insect pin.
- Videos were filmed at a rate of 80,000 to 100,000 frames per second (the three A. horridus
- 17 filmed with the Miro eX4 were filmed at 58,823 fps). We recorded between three and five
- individuals for each species, and between four and twenty strikes per individual (Table 1). After
- recording, each ant was preserved in a -20°C freezer for at least 24 hours. Within one week of
- 20 filming, the mass of each ant was measured with a UMX2 ultra microbalance (Mettler Toledo,
- 21 Columbus, OH). Ants were then imaged using a Leica M205 C stereo microscope (467 nm
- resolution) attached to a five-megapixel Leica DFC 425 digital microscope camera, and the head
- width, head length, and mandible lengths measured. The mandibles were then removed and
- 24 weighed individually using the microbalance.
- To digitize mandibles strikes, we tracked the tips of each mandible in each video in
- 26 MATLAB (2014a) using a custom script taken from Spagna et al. (2008). The cumulative
- 27 displacement of the mandibles was then calculated trigonometrically from the x-y coordinates of
- 28 the mandible tips generated by this script. These values were then exported to R (ver3.3.3) where
- 29 the cumulative angular displacement was smoothed using a quintic spline via the penalized
- 30 smoothing splines package (pspline). Instantaneous rotational velocity and rotational acceleration
- were calculated by taking the first and second derivatives of displacement, respectively, and used

- 2 were calculated by multiplying the maximum rotational velocity and acceleration by the length
- 3 of the mandible. Rotational kinetic energy (K) was calculated by modeling each mandible as a
- 4 thin rod of uniform density rotating about one end, using the equation:

$$K = \frac{1}{6}mr^2w^2$$

- 5 Where m is the mandible mass, r is the mandible length, and w is the maximum rotational
- 6 velocity. Maximum rotational kinetic energy and strike duration were used to estimate maximum
- 7 power output (P) for each strike, using the equation:

$$P = \frac{K}{tm_a}$$

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- 8 Where t is the duration of the strike and m_a is the estimated mass of the mandible closer muscle.
- 9 This estimate assumes a combined mandible closer muscle mass equal to one fourth that of the
- ant's body mass based on the relationship between mandible closer's size and body mass in other
- odontomachine species (Larabee unpublished data) and is a conservative estimate of the
- maximum power output of the mechanism. Our goal in calculating power output is not to obtain
- absolute measurements for each species, but rather estimates that will allow us to determine if
- they are using a power amplification mechanism. Time delay between the release of each
- mandible during strikes was calculated by subtracting the frame number of the first mandible's
- release from the frame number of the second, and dividing the result by the frame rate.

MicroCT

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- To examine the internal arrangement of head musculature in *Anochetus*, one individual of A.
- 20 horridus was examined using X-ray microtomography (microCT). The ant was fixed in alcoholic
- 21 Bouin's solution (Sigma Aldrich Corporation, Steinheim, Germany) for 24 hours, after which it
- was washed in 70% ethanol and dehydrated through an ethanol series (at least 20 min each at
- 23 70%, 80%, 90%, 95%, and two changes of 100%). The ant was stained overnight in I2E (2%)
- iodine w/v in 100% ethanol) to increase contrast of muscle tissue and then washed in 100%
- 25 ethanol. To preserve microstructure of soft tissue, the specimen was critical point dried
- 26 (AutoSamdri-931.GL Supercritical Point Dryer, Tousimis Research Corporation, Rockville,
- MD), and stored in a desiccator for approximately 24 hours before scanning. The head of the ant
- was placed in an appropriately sized pipette tip which was sealed with cotton and scanned using

- an Xradia MicroXCT-400 scanner (Carl Zeiss, Oberkochen, Germany) set at 25kV voltage and
- 2 5W power with an exposure time of three seconds. 1441 images were taken at different rotations
- 3 spanning 180°. Tomographic reconstruction was performed in Xradia XMReconstructor 8.1.
- 4 Volume renderings and surface models were produced in Amira 5.6.0 (FEI, Hillsboro, OR) and
- 5 used to calculate the relative volumes that the mandible abductor, adductor, and trigger muscles
- 6 occupy in the head capsule.

8 Statistics

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- 9 To compare the kinematic properties of mandible strikes between species, only the fastest of the
- 10 left and right mandible were used for each strike. Kinematic properties of mandible strikes were
- compared between species using an ANOVA and post hoc Tukey HSD test with strikes treated
- as a random effect nested within individuals. All stats were performed in R version 3.3.3 (CRAN
- 13 Studios) using the package nlme (Pinheiro *et al.*, 2017).

Results

- The species of *Anochetus* examined in this study exhibit a five-fold difference in body size
- between the smallest and largest species (Table 1; Figure 1). The mass-specific power of
- Anochetus strikes ranged from 4.3×10^4 to 1.9×10^5 W/kg, with all species producing strikes that
- 19 have power output values far above those possible through direct muscle contraction alone
- 20 (Josephson, 1993). The kinematic profiles of *Anochetus targionii* and *A. paripungens* resemble
- 21 those of O. ruginodis, the smaller of the two Odontomachus species examined in this study
- 22 (Figure 2, black curves). Anochetus horridus and A. emarginatus, in contrast, have different
- 23 strike kinematic profiles from the other *Anochetus* species examined (Figure 2).
- Mean strike durations for A. horridus (mean \pm SD: 0.251 ± 0.11 ms) and A. emarginatus
- 25 $(0.22 \pm 0.08 \text{ms})$ are longer than A. targionii $(0.101 \pm 0.04 \text{ms})$ and A. paripungens $(0.103 \pm 0.04 \text{ms})$
- 26 0.04ms) and are similar to strike durations recorded for the larger O. ruginodis (0.19 \pm 0.09ms)
- and O. chelifer $(0.24 \pm 0.09 \text{ms}; \text{ Figure 3A})$. Anotherus targionii and A. paripungens both have
- similar maximum rotational velocities (37,000 \pm 3,000rad/s and 36,500 \pm 3,000rad/s,
- 29 respectively, rounded to the nearest five hundred) that are significantly higher than those seen in
- 30 *Odontomachus* (31,500 \pm 6,000rad/s for *O. ruginodis* and 17,000 \pm 3,500rad/s for *O. chelifer*;
- 31 Figure 3B). Anochetus horridus and A. emarginatus have maximum rotational velocity values

- that are similar to each other and are the slowest of all species examined in this study (14,500 \pm
- 3,000rad/s and $12,500 \pm 2,500$ rad/s, respectively). Maximum rotational velocity is attained at
- 3 approximately the same time for all species, but A. horridus and A. emarginatus accelerate and
- 4 decelerate at a slower rate than other *Anochetus* and *O. ruginodis* (Figure 2B, C). Mean linear
- velocity was lowest in the mandible strikes of A. horridus (19.9 \pm 4.3m/s) and A. emarginatus
- 6 (23.6 \pm 4.2m/s) and highest in O. chelifer (40.0 \pm 7.5m/s), with all other species obtaining
- intermediate maximum velocities (A. targionii: 29.4 ± 2.4 , A. paripingens: 29.0 ± 2.7 m/s, O.
- 8 ruginodis: 33.7 ± 6.3 ; Figure 3C).
- Maximum rotational acceleration was lowest in A. horridus $(2.6 \times 10^8 \pm 8.4 \times 10^7 \text{rad/s}^2)$ and
- 10 A. emarginatus $(1.8 \times 10^8 \pm 4.5 \times 10^7 \text{ rad/s}^2)$, both of which were similar in value to the larger O.
- 11 chelifer $(2.8 \times 10^8 \pm 8.1 \times 10^7 \text{ rad/s}^2)$. All other species of Anochetus and Odontomachus examined
- obtained peak rotational accelerations two to three times that of A. horridus and A. emarginatus
- 13 (A. targionii: $8.8 \times 10^8 \pm 1.1 \times 10^8 \text{ rad/s}^2$, A. paripungens: $8.5 \times 10^8 \pm 9.6 \times 10^7 \text{ rad/s}^2$, O. ruginodis:
- $7.0 \times 10^8 \pm 1.7 \times 10^8 \text{ rad/s}^2$; Figure 3D). Linear acceleration followed the same pattern, with A.
- horridus and A. emarginatus reaching the lowest peak linear acceleration $(3.5 \times 10^5 \pm 1.1 \times 10^5 \text{m/s}^2)$
- and $3.5 \times 10^5 \pm 8.3 \times 10^4 \text{m/s}^2$, respectively) and all other species reaching peak linear accelerations
- approximately two times greater than A. horridus and A. emarginatus (A. targionii: $7.1 \times 10^5 \pm$
- 18 8.9x10⁴m/s², A. paripungens: $6.8x10^5 \pm 9.0x10^4$ m/s², O. ruginodis: $7.5x10^5 \pm 1.9x10^5$ m/s², O.
- 19 *chelifer*: $6.5 \times 10^5 \pm 1.8 \times 10^5 \text{m/s}^2$; Figure 3E).
- 20 Maximum kinetic energy is positively associated with body size (Figure 3F). The
- smallest three *Anochetus* species had the lowest kinetic energy values (A. targionii: 2.0 ± 0.31
- μ J, A. horridus: $0.93 \pm 0.35 \mu$ J, A. paripungens: $2.4 \pm 0.49 \mu$ J), while A. emarginatus and O.
- ruginodis, the two medium sized species, had intermediate kinetic energy values (A.
- emarginatus: $7.1 \pm 2.3 \,\mu\text{J}$, O. ruginodis: $10 \pm 3.4 \,\mu\text{J}$). Odontomachus chelifer, the largest
- species, had the most energetic strike of all species examined ($100 \pm 35 \mu J$).
- In most strikes examined the mandibles were not released simultaneously (*A. horridus*:
- 40 of 41 strikes; A. emarginatus: 18 of 22 strikes; A. targionii: 14 of 17 strikes; A. paripungens:
- 28 of 32 strikes); lag times between each mandible starting its acceleration averaged from 0.03 to
- 29 0.12ms among species (Figure 4). This variation was significant (nested ANOVA, F=6.7, df=5,
- p<0.001) and paired comparisons revealed that A. horridus had longer lag times between
- mandibles than A. targionii and A. paripungens (Tukey HSD, p<0.001; other paired comparisons

- p>0.05). Maximum rotational and linear velocity did not differ significantly between the fastest
- 2 and slowest mandible within a strike for any species except A. paripungens (paired t-test,
- 3 rotational velocity: t=6.9, df=62, p<0.0001; linear velocity: t=4.3, df=62, p<0.0001). In A.
- 4 targionii, the first mandible to be released tended to reach a higher maximum rotational velocity
- than the second (exact binomial test, number of successes =13, number of trials =14, p=0.001).
- 6 This was not the case for A. paripungens (exact binomial test, number of successes=9, number of
- 7 trials=28, p=0.08), A horridus (exact binomial test, number of successes=14, number of
- 8 trials=31, p=0.72), or A. emarginatus (exact binomial test, number of successes=5, number of
- 9 trial=13, p=0.58). In these species, there is no relationship between which mandible is released
- 10 first and which obtains a higher velocity. As in *Odontomachus bauri* (Patek et al., 2006),
- 11 Anochetus mandibles begin to decelerate before crossing the midline (Figure 4B, C).
- The microCT scan of A. horridus revealed the mandible closer (adductor) muscles (AdM)
- occupy most of the ventral and posterior sections of the head capsule and account for 20% of the
- total head capsule volume (Figure 5). Contraction of these muscle when the mandibles are
- locked open likely deforms the mandible apodemes, allowing elastic energy to be stored prior to
- a strike. The trigger muscles (fAdM) are specialized sections of the mandible closer muscle that
- consist of a small group of muscle fibers inserting on the mandible adductor apodeme
- ventrolaterally and originating at the ventrolateral side of the postgenal section of the head
- capsule. They occupy 0.17% of the head capsule volume and account for 0.9% of the volume of
- 20 the mandible adductor muscle. These fibers are likely fast twitch muscle fibers based on previous
- studies of this muscle in other species of trap-jaw ants, and their contraction rotates the
- 22 mandibles out of their locked position and allows the mandibles to swing shut when the elastic
- energy stored in the apodemes is released. The mandible opener (abductor) muscles (AbM) are
- much smaller than their closer counterparts, occupying 2.5% of the total head capsule volume.
- The mandible abductor muscles insert near the base of the mandible and attach to the medial
- portion of the postgena in between the closer muscles. Contraction of these muscles opens the
- 27 mandibles and, if contracted far enough, locks the mandibles into the open position.

Discussion

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Influence of body size and morphology on strike performance

- 1 Our estimates of mass specific power output for each species exceed those possible through
- 2 muscle contraction alone (Josephson, 1993), confirming that *Anochetus* mandible strikes are
- 3 indeed power amplified. All species followed a pattern of increasingly energetic strikes with
- 4 larger body mass. This relationship is likely driven primarily by the increase in mandible mass as
- 5 body size increases. *Anochetus targionii* and *A. paripungens* mandible strikes closely resemble
- 6 those of *Odontomachus ruginodis*, although they consistently outperform *O. ruginodis* in many
- 7 aspects of their strike kinematics, on average obtaining higher peak rotational velocities and
- 8 accelerations. As both species are considerably smaller than O. ruginodis, these results are
- 9 consistent with the relationship between strike performance and body size found in
- 10 Odontomachus (Spagna et al., 2008). In contrast, A. horridus and A. emarginatus (both from the
- 11 *emarginatus* species group) have similar mandible strikes that consistently underperform other
- 12 Anochetus and Odontomachus species, on average obtaining lower peak velocities and
- accelerations. This relationship cannot be explained by relative differences in body size, as the
- body mass of A. horridus is intermediate between A. targionii and A. paripungens, and one fifth
- that of A. emarginatus. Other factors, such as similar head and mandible morphology, differing
- life history strategies, or evolutionary constraints (Larabee et al., 2016) may partially explain
- differences in strike performance of A. horridus and A. emarginatus relative to other Anochetus
- 18 species.
- Both A. horridus and A. emarginatus have relatively long, thin mandibles with respect to
- 20 their head width compared to the other *Anochetus* species. The average mandible length to head
- width ratio is 1.32 and 1.12 for A. horridus and A. emarginatus, respectively, compared to 0.74
- and 0.69 for A. targionii and A. paripungens. These differences in relative mandible length,
- 23 however, do not translate to differences in relative mass; the average mandible mass to body
- mass ratio is 0.011 and 0.012 for A. horridus and A. emarginatus compared to 0.011 and 0.010
- for A. targionii and A. paripungens. Having more elongate mandibles increases the minimum
- 26 distance needed between the ant and its prey for a strike to be successful, possibly increasing the
- 27 chances of successfully capturing prey. The reduced strike performance seen in A. horridus and
- 28 A. emarginatus may be an adaptation to prevent their mandibles from fracturing under the
- 29 increased stresses of a more forceful strike. The reduced strike performance in this species group
- 30 could therefore be the result of a trade-off between the probability of successfully capturing prey

and the amount of internal stress the mandible experiences when biting, an idea that warrants future study.

Influence of physiology on strike performance

Differences in muscle geometry and composition might explain differences in *A. horridus* and *A. emarginatus* strike performance (see for example David *et al.*, 2016). However, the muscle arrangement within the head capsule of *A. horridus* we report here is similar to that of *A. graeffei* as described by Gronenberg & Ehmer (1996). In ants, muscle attachment angle and the proportion of mandible closer muscle consisting of fast and slow twitch muscle fibers both influence bite performance (Gronenberg *et al.*, 1997; Paul & Gronenberg, 1999). *Anochetus* species do not possess fast twitch fibers in their mandible closer muscle and rely on stored elastic energy (likely in the muscle apodeme or cuticle of the head) to provide the speed needed for their strikes (Gronenberg & Ehmer, 1996). Differences in strike kinematics are thus likely not due to underlying physiological differences in mandible closer muscle fibers between species, and likely reflect differences in the material properties of the elastic energy storing component of the mechanism. Determining the specific structures responsible for elastic energy storage within the head capsule of *Odontomachus* and *Anochetus* prior to mandible release and examining species level differences in the material properties of those structures may help to explain performance differences in future studies.

The venom potency of trap-jaw ants is another aspect that may influence the mandible strike performance. Both mandibles and venom are important features for prey capture and one may be negatively correlated to the other. This is well illustrated in scorpions where scorpions with more potent venom possess relatively slender claws (Van der Meijden *et al.*, 2010). Although little data on the potency and the toxicity of ant venoms are available, both *A. horridus* and *A. emarginatus* have unusual venom compositions that vastly contrast with venoms of *Odontomachus* and other ponerine ants (Touchard *et al.*, 2015). Furthermore, the investigation of the *A. emarginatus* venom resulted in the discovery of a novel family of neurotoxins that reversibly paralyses insects (Touchard *et al.*, 2016). Nevertheless, further venom investigations extended to other *Anochetus* species are needed to support this hypothesis.

All four *Anochetus* exhibited a lag between the release of the first and second mandibles during a strike. These lag times were similar to those observed in both *Odontomachus* species, and there was no difference in performance between the first and second mandible to be released

with the exception of A. targionii, in which the first mandible to be released consistently reached 1 2 a higher maximum rotational velocity compared to the second. This contrasts with the findings 3 of Patek et al. (2006), who found that in O. bauri the second mandible to be released usually 4 reached a higher maximum velocity. A delay between mandibles in their release may be the result of the time it takes to transmit a signal from sensory neurons to the corresponding motor 5 neurons (Patek et al., 2006). In A. graeffei, the sensory neurons attached to trigger hairs do not 6 cross over to the opposing side of the subesophogeal ganglion like they do in *Odontomachus* 7 (Gronenberg & Tautz, 1994; Gronenberg & Ehmer, 1996), suggesting that Anochetus should 8 have longer lag times compared to its sister genus. Our results are not consistent with this 9 pattern, possibly because the difference in lag time is so minute that we were not able to detect it 10 at the frame rates used in this study. It is also possible that this neuronal configuration is 11 12 restricted to certain species like A. graeffei, and in the species examined in this study crossing over of the sensory trigger neurons does occur in a similar manner to *Odontomachus*. Future 13 14 studies using histological methods to examine how neurophysiology varies between species and examining the performance of A. graeffei mandible strikes will be able to answer this question. 15 16 Lag time between mandible release is likely restricted to ponerine trap-jaw ants; in many myrmicine trap-jaw ants (Daceton, Orectognathus, Epopostruma, and Strumigenys) the labrum 17 18 is modified to act as the latch, allowing the mandibles to be released simultaneously (Gronenberg, 1996b). 19 20 In all species examined, both mandibles begin to decelerate before crossing the midline, similar to *Odontomachus* mandible strikes (Patek et al., 2006). This premature deceleration is 21 22 thought to be adaptive as it would prevent the mandibles from crashing into each other at full force if the ant misses its target. Examining the kinematics of more phylogenetically independent 23 24 trap-jaw mechanisms (e.g. in other ant genera) will determine if this deceleration is a universal characteristic of trap-jaw mandibles to reduce mandible wear or if it is restricted to certain taxa. 25 26 Most myrmicine trap-jaw ant genera, for example, are smaller bodied and likely generate less force than ponerine trap-jaw ants which have more massive mandibles. Preventing mandibles 27 28 from colliding at full force may be less of an issue in smaller species. 29 Gronenberg & Ehmer (1996) described the trap-jaw mechanism in A. graeffei, noting that the morphology, musculature, and neurophysiology of the mechanism are generally similar to 30 Odontomachus but differ in a few key characteristics. Anochetus graeffei has a trigger muscle 31

- 1 composed of fibers that attached indirectly to the mandible apodeme via short, thin filaments,
- 2 while in *Odontomachus* the trigger muscle fibers all attach to the apodeme directly (Gronenberg
- 3 & Ehmer, 1996). The mandible closer muscle of A. graeffei also contains more filament-attached
- 4 muscle fibers compared to the closer muscle in *Odontomachus* (Gronenberg & Ehmer, 1996).
- 5 This interspecific variation in physiology may account for some of the differences we see in
- 6 kinematics between *Anochetus* and *Odontomachus*. The microCT scan of *A. horridus* suggests
- 7 that muscle arrangement is likely consistent across species of *Anochetus* as muscle arrangement
- 8 in A. horridus closely resembles that of A. graeffei at the level of detail discernable via MicroCT
- 9 scans. More detailed studies comparing head capsule musculature and innervation at the cellular
- 10 level are needed to confirm this.

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Influence of phylogeny on strike performance

- The confounding effect of shared evolutionary relatedness may also partially explain the
- strike performance patterns we observed. *Anochetus horridus* and *A. emarginatus* are closely
- related, both being members of the *emarginatus* species group (Larabee *et al.*, 2016). In contrast,
- A. targionii and A. paripungens originate from the phylogenetically and geographically distinct
- mayri and rectangularis species groups (distributed throughout the new and old world tropics
- 17 respectively; Larabee et al., 2016). The strong similarities in their strike performance suggests
- that morphology and body size are a large component of what determines strike performance in
- this genus. However, the scope of this study does not allow for large scale comparisons of the
- 20 relative roles of phylogeny and morphology on strike performance.

Conlcusion

- Our results suggest that phylogeny, physiology, morphology and natural history of trap-
- 23 jaw ant species may each play an important role in determining strike performance, although
- 24 understanding the relative role of each will require describing the kinematics of a larger number
- of species in an explicit phylogenetic context. The repeated evolution of power amplified
- 26 mandibles in "trap-jaw" ants has been implicated in increased diversification rates and lineage
- persistence (Pie & Tscha, 2009; Moreau & Bell, 2013; Larabee & Suarez, 2014), although this
- 28 idea has not yet been tested empirically. Future work should focus on describing the strike
- 29 performance of additional *Odontomachus* and *Anochetus* species, and combine this data with
- 30 phylogenetic (Larabee et al., 2016), natural history (Brown, 1976; Brown, 1978), and
- 31 morphological information to examine the relative importance of these factors on strike

- 1 performance. Future studies that study the performance of other independent evolutions of trap-
- 2 jaw mechanisms, such as those in the subfamily Myrmecinae, will also shed light on which
- 3 kinematic properties of mandible strikes, if any, are universal to all trap-jaw ants.

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- 19 71.403850°W) in August of 2013; A. horridus was collected from under a log and a colony of A.
- 20 emarginatus was collected from the roots of epiphytes at the Nouragues National Nature

from leaf litter at the Villa Carmon Biological Research station in Peru (12.89474°S

- 21 Reserve, French Guiana (3.982411°S 52.563872°W) in March of 2016; A. paripungens was
- collected from the soil in Humpty Doo, NT, Australia (12°34'21.37"S 131° 5'2.43"E) in April of
- 23 2010; O. chelifer was collected from Viçosa, Minas Gerais, Brazil (20°45'28"S 42°51'46"W) in
- October of 2014; O. ruginodis was collected in October of 2005 from Archbold Biological
- 25 Station in Florida (27°11'5.61"N 81°20'19.64"W).

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- 27 **Figure 1.** Representative images of study species displaying the morphological and size
- variation present within *Anochetus* and *Odontomachus*. Images are to scale. A: *Anochetus*
- 29 targionii. B: Anochetus horridus. C: Anochetus paripungens. D: Anochetus emarginatus. E:
- 30 Odontomachus chelifer. F: Odontomachus ruginodis. All images are from Antweb.org.

- 1 **Figure 2.** Representative kinematic profiles of *Anochetus* mandible strikes. A: Angular
- 2 displacement versus time. Points represent raw cumulative displacement data, curves are the
- 3 spline fitted to those points. B: Angular velocity versus time. C: Angular acceleration versus
- 4 time. D: representative frames of the A. targionii mandible strike described in A-C. E:
- 5 representative frames from the video of the A. emarginatus mandible strike described in A-C.
- 6 For panels A-C, species are indicated by color: Blue: A. targionii. Red: A. horridus. Purple: A.
- 7 *emarginatus*. Black: *Odontomachus ruginodis*. *Odontomachus chelifer* closely resembles *O*.
- 8 ruginodis and was not included in these graphs. Numbers between D and E indicate the time in
- 9 milliseconds relative to the start of the mandible strike. Strikes depicted here are shown in
- 10 Supplemental Videos 1-4.
- 11 **Figure 3.** Comparative mandible strike kinematics for *Anochetus* and *Odontomachus* species
- included in this study. Values shown are means of the maximum values for the left and right
- mandibles of each strike. Species are arranged by body mass, with the smallest species on the left
- and the largest species on the right. Error bars denote one standard deviation from the mean. All
- variables examined differed significantly between species (Nested ANOVA, df=5, p<0.05). Bars
- that share letters at their bases are not statistically distinguishable from one another based on a
- Tukey HSD test at the p=0.05 level. A: strike duration; B: maximum rotational velocity; C:
- maximum linear velocity; D: maximum rotational acceleration; E: maximum linear acceleration,
- F: maximum kinetic energy (O. chelifer excluded from graph due to large difference from other
- 20 species).
- Figure 4. Delay in mandible release (the time difference between when the first and second
- 22 mandible begin to move) is present in all *Anochetus* spp. examined. A-C: graphs displaying
- 23 kinematic profiles of the left and right mandibles during a representative A. targionii strike (the
- same strike depicted in Figure 2). Points represent the raw displacement data used to calculate
- 25 the spline. Solid lines represent the left mandible, dotted lines represent the right mandible. A:
- angular displacement vs time; B: angular velocity vs time; C: angular acceleration vs time. D:
- 27 Box and whisker plot depicting the time delay in mandible release between the left and right
- 28 mandibles for all mandible strikes examined.
- 29 **Figure 5.** Reconstructed model of *Anochetus horridus* head capsule from a micro-CT scan. A:
- 30 2D coronal section of head capsule. B: dorsal view, with the left side of the head capsule
- 31 removed to show the arrangement of muscles relevant to the trap-jaw mechanism and the right

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side showing the external features of the head. C: Lateral view cut away to display the same 1 2 structures labeled in A. AdM: mandible adductor muscle, AbM: mandible abductor muscle, 3 fAdM: fast twitch adductor muscle (trigger muscle), M: mandible and mandible apodeme. Voxel size: $2.0 \, \mu \text{m}^3$. 4 **Supporting Information** 5 **Supplemental Video 1:** representative mandible strike of *Anochetus targionii*. The strike is the 6 same one depicted in figures 2A-D and 4A-C. The video was filmed at 80,000 frames per 7 8 second. 9 Supplemental Video 2: Representative mandible strike of Anochetus horridus. The strike is the 10 same one depicted in figure 2A-C. The video was filmed at 100,000 frames per second. 11 12 **Supplemental Video 3:** Representative mandible strike of *Anochetus paripungens*. The strike is 13 the same one depicted in figure 2A-C. The video was filmed at 88,888 frames per second. 14 15 Supplemental Video 4: Representative mandible strike of Anochetus emarginatus. The strike is 16

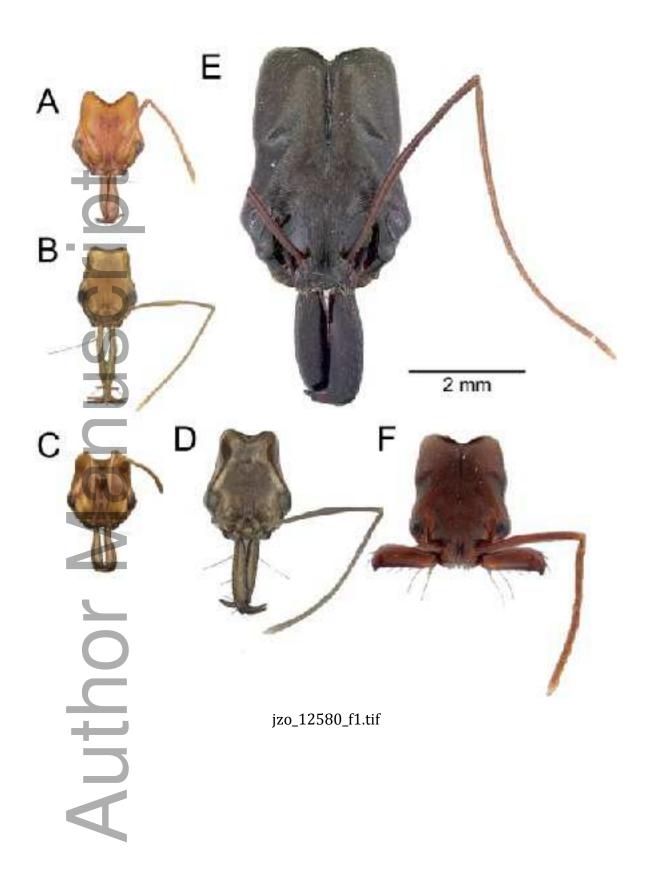
the same one depicted in figure 2A-C, E. The video was filmed at 80,000 frames per second.

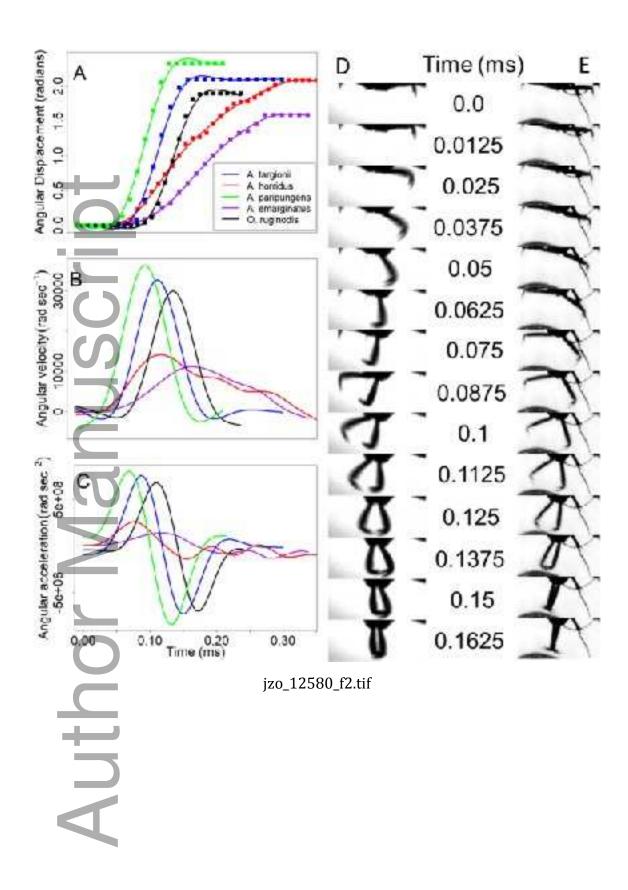
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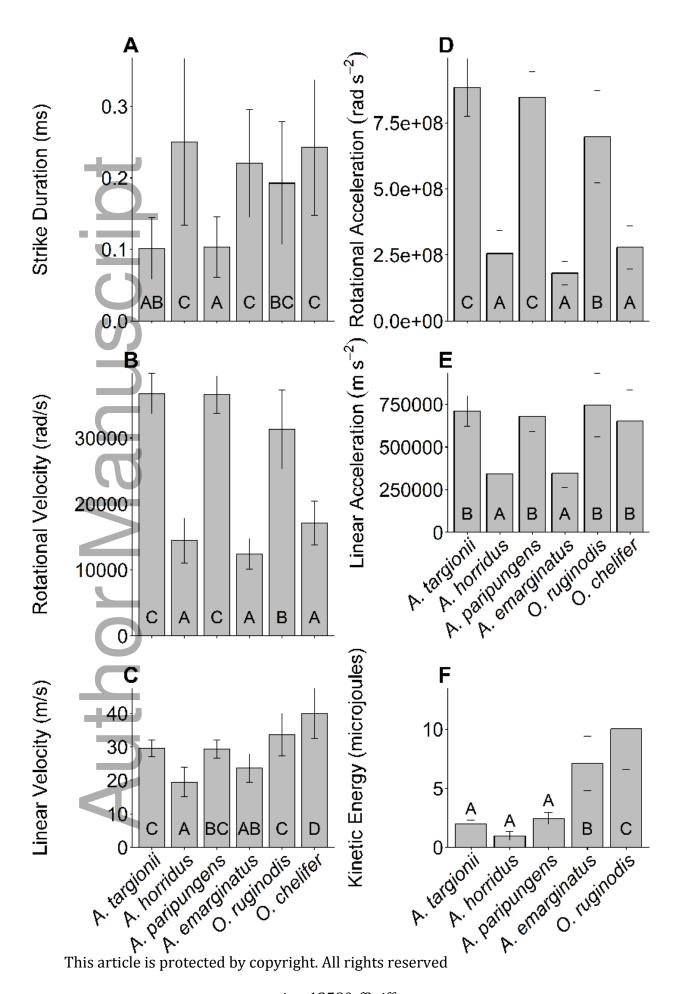
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Species	Locality	n	i	Head width (mm)	Body mass (mg)	Mandible length (mm)	Mandible mass (μg)	Estimated closer muscle mass (mg)
A. targionii	Peru	17	4	1.07 ± 0.013	1.17 ± 0.001	0.80 ± 0.01	13.65 ± 0.64	0.29 ± 0.02
A. horridus	French Guiana	41	5	1.03 ± 0.015	1.39 ± 0.09	1.33 ± 0.03	12.99 ± 6.3	0.35 ± 0.02
A. paripungens	Australia	32	5	1.17 ± 0.013	1.62 ± 0.1	0.81 ± 0.04	16.69 ± 1.9	0.41 ± 0.03
A. emarginatus	French Guiana	22	3	1.72 ± 0.03	5.85 ± 0.31	1.93 ± 0.1	72.72 ± 5.7	1.46 ± 0.08
O. ruginodis	Florida, USA	15	5	1.62 ± 0.1	5.66 ± 0.05	1.08 ± 0.03	52.33 ± 6.9	1.42 ± 0.13
O. chelifer	Brazil	15	3	2.68 ± 0.17	30.1 ± 5.6	2.34 ± 0.13	366.33 ± 93	7.53 ± 1.4
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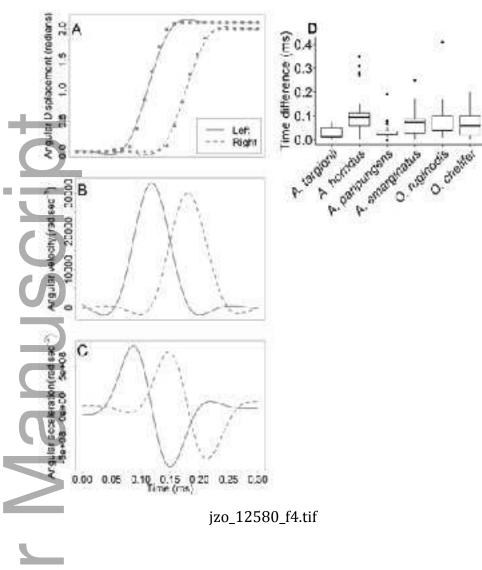
Table 1. Summary of *Anochetus* and *Odontomachus* measurements

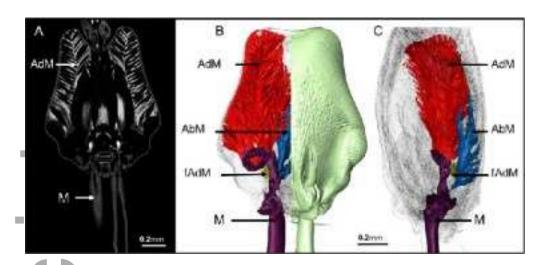






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